# Stereospecific Acetolysis of threo- and erythro-2-Aryl-2-[1-(methylthio)ethyl]oxiranes

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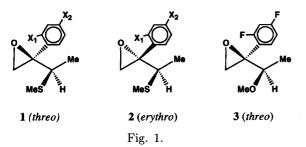
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Threo-2-aryl-2-[1-(methylthio)ethyl]oxiranes ( $1\mathbf{a}$ — $\mathbf{d}$ ) were reacted with acetic acid to give two types of unexpected ring-opening products, threo-1-acetoxy-2-aryl-3-methylthio-2-butanols ( $4\mathbf{a}$ — $\mathbf{d}$ ) and mono-O-acetyl derivatives of threo-2-aryl-2-methylthio-1,3-butanediol ( $5\mathbf{a}$ — $\mathbf{d}$  and  $6\mathbf{a}$ — $\mathbf{d}$ ). On the other hand, erythro-2-aryl-2-[1-(methylthio)ethyl]oxiranes ( $2\mathbf{a}$ — $\mathbf{d}$ ) gave erythro-1-acetoxy-2-aryl-3-methylthio-2-butanols ( $7\mathbf{a}$ — $\mathbf{d}$ ) and mono-O-acetyl derivatives of erythro-2-aryl-2-methylthio-1,3-butanediol ( $8\mathbf{a}$ — $\mathbf{d}$  and  $9\mathbf{a}$ — $\mathbf{d}$ ). It is suggested that these products are generated from thiiranium ionic intermediates, because they all completely retain the relative configuration of the starting oxiranes.

 $\alpha$ -Substituted styrene oxides are important intermediates for the preparation of potent antifungal compounds.<sup>1-3)</sup> We previously reported the synthesis of *threo*- and *erythro*-2-aryl-2-(1-substituted ethyl)oxiranes (1-3) in a diastereoselective manner (Fig. 1).<sup>2)</sup> In the course of our study on the reactivities of these oxiranes, we observed that oxiranes 1 and 2 gave two types of unexpected ring-opening products upon acetolysis. We herein describe the details of this reaction and the proposed reaction mechanisms.

## Results and Discussion

threo-2-Aryl-2-[1-(methylthio)ethyl]oxiranes (1a—d) were treated with acetic acid in dichloromethane at 40 °C to give threo-1-acetoxy-2-aryl-3-methylthio-2-butanols (4a—d) and unexpected rearrangement products, mono-O-acetyl derivatives of threo-2-aryl-2-methylthio-1,3-butanediol (5a—d and 6a—d) as shown in Scheme 1. On the other hand, erythro-2-aryl-2-[1-(methylthio)ethyl]oxiranes (2a—d) gave erythro-1-acetoxy-2-aryl-3-methylthio-2-butanols (7a—d) and mono-O-acetyl derivatives of erythro-2-aryl-2-methylthio-1,3-butanediol (8a—d and 9a—d). Interestingly, threo-oxiranes 1a—d gave only threo-products, and erythro-oxi-



ranes **2a**—**d** gave only *erythro*-products. Moreover, neither *threo*- nor *erythro*-2-acetoxy-2-(2,4-difluorophenyl)-3-methylthio-1-butanol (**10** or **11**) were obtained at all, even though they were expected as normal ring-opening products under acidic conditions. The yield and product ratio in the acetolysis of *threo*-oxiranes **1a**—**d** and *erythro*-oxiranes **2a**—**d** are shown in Tables 1 and 2. In all cases, products **4a**—**d** were easily separated from the reaction mixture by silica-gel column chromatography, but **5** and **6** couldn't be. Thus, the product ratio was

Table 1. Acetilysis of threo-Epoxide 1a—d

	Substituent		Total yield <sup>a)</sup>	Product ratio <sup>b)</sup>
	$\overline{X_1}$	$X_2$	%	4:5:6
1a	F	F	98.6	94:2:4
1b	H	$\mathbf{F}$	95.1	96:2:2
1c	H	H	96.1	94:3:3
1d	H	Me	92.1	94:3:3

a) Isolated yield of product 4, 5, and 6. b) Ratio of 4 vs. (5+6) was determined by isolated yield. Ratio of 5 vs. 6 was determinated by <sup>1</sup>H NMR measurement.

Table 2. Acetilysis of erythro-Epoxide 2a—d

	Substituent		Total yield <sup>a)</sup>	Product ratio <sup>b)</sup>	
	$\overline{X_1}$	$X_2$	%	7:8:9	
2a	F	F	87.5	24:57:19	
2b	H	$\mathbf{F}$	82.1	37:25:38	
2c	H	H	82.4	37:24:39	
2d	H	Me	81.2	58:17:25	

a) Isolated yield of product 7, 8, and 9. b) Ratio of 7 vs. (8+9) was determinated by isolated yield. Ratio of 8

vs. 9 was determinated by <sup>1</sup>H NMR measurement.

i) K<sub>2</sub>CO<sub>3</sub> (2.0 eq.), in MeOH, room temp., 3 h, 92.9 % yield.

ii) 4-chloroperbenzoic acid (3.0 eq.), in CH<sub>2</sub>Cl<sub>2</sub>, room temp., 1d., 81.6 % yield.

# Scheme 2.

Fig. 2.

determined from the isolated yields of 4 vs. 5 and 6, and from the NMR peak area ratio of 5 vs. 6. The product ratio of 7:8:9 was determined by the same method.

In the reaction of threo-oxiranes 1a—d with acetic acid, 4a—d were given as main products in high yields

as shown in Table 1. The ratio of products (4:5:6) was quite independent of the substituent pattern on the phenyl ring. On the other hand, in the reaction of *eryth-ro*-oxiranes 2a—d with acetic acid, 8a—d and 9a—d were given in considerable yields as shown in Table 2. In

Scheme 3.

addition, the yield of 7 became lower, as the electron-withdrawing factor of the substituents on the phenyl ring increased. In contrast, the corresponding 3-methoxy analog, 3, was quite stable under the same reaction conditions, and didn't give any ring-opening product. The reactivity of the oxiranes is obviously enhanced by the methylthio group.

then 15 % aq. KOH

In this connection, the relative configuration of 8a and 9a was confirmed to be *erythro* by X-ray crystallographic analysis of crystalline sulfone 13, which was given from the mixture of 8a and 9a in 2 steps as shown in Scheme 2. The 3:1 mixture of 8a and 9a was hydrolyzed to give diol 12a in a 92.2% yield, which was

oxidized by 4-chloroperbenzoic acid to afford sulfone 13 as crystals after recrystallization from dichloromethane.

The proposed reaction mechanism can be summarized in Figs. 2 and 3. From the fact that all the products retain the relative configurations of the starting oxiranes, the mode of the reaction isn't assumed to be  $S_N1$  via a benzyl cationic intermediate, but rather  $S_N2$  via the thiiranium ionic intermediate. The nucleophilic attack of the acetoxyl group on the thiiranium ionic intermediate should take place with complete inversion. Such a thiiranium ionic intermediate will give two types of adducts, since it has two possible positions for nucleophilic attack. When the C-2 position of the intermedi

ate suffers the nucleophilic attack, 2-acetoxy-1-ols (10 or 11) should be produced (path A). However, we could not detect such compounds, because they are probably transformed to the corresponding 1-acetoxy-2-ols (4 or 7) at once via acyl rearrangement.<sup>4)</sup> In the meantime, when the C-3 position of the intermediate is the target of the nucleophilic attack, sulfur-rearranged product 5 or 8 is yielded (path B). Furthermore, products 6 and 9 are obtained by rearrangement of acyl group on 5 and 8, respectively.<sup>4)</sup>

The difference in the regional regional regional regions are the regional region of the regional regions. philic attack of the acetoxyl group on the thiiranium ionic intermediate between the three-series and eruthroseries can be explained as follows. three-Isomers 1a—d prefer path A, because it is easy for thiiranium ionic intermediates generated from them to undergo nucleophilic attack at the more electrophilic C-2 position. Since the phenyl group and the methyl group on these intermediates are placed in trans geometry, the C-2 position of the intermediates should not be so bulky (Fig. 2). On the other hand, in the case of thiiranium ionic intermediates generated from erythro-isomers **2a**—**d**, the *cis*-methyl group prevents the rotation of the phenyl ring, and this brings steric hindrance to the C-2 position. Thus, the nucleophilic attack occurs at the C-3 position to give rearranged products 8 and **9** in considerable yields (Fig. 3).

With respect to the preparation of the starting materials, threo-epoxides 1a—d, free from their erythroisomers, were prepared as shown in Scheme 3.5 threo-Rich epoxide 1a (threo/erythro=5:1) afforded by a previously reported method, 2 was reacted with acetic acid similar to the method described above. After purification, major product 4a was hydrolyzed to give threodiol 14a. Diol 14a was treated with methanesulfonyl chloride and triethylamine followed by treatment with aqueous KOH solution to regenerate threo-epoxide 1a free from its erythro-isomer. Other threo-epoxides 1b, 1c, and 1d were prepared by the same method.

Although a number of examples of neighboring sulfur participation have been studied, <sup>6)</sup> the formation of thiiranium ions from 2,3-epoxy sulfides has not been reported to any significant degree. <sup>7)</sup> Moreover, diols derived from **1a**—**d** by hydrolysis are also useful for synthesis of antifungal agents. <sup>8)</sup> Further studies on applications of this reaction will be reported in the near future.

## Experimental

Melting points were determined on Thomas–Hoover capillary melting point apparatus and were uncorrected. Infrared spectra (IR) were recorded on a JASCO A-102 IR spectrometer. Proton magnetic resonance spectra ( $^{1}$ H NMR) were obtained on a JEOL JNM-GX270 spectrometer (270 MHz) in the designated solvent using tetramethylsilane as an internal standard ( $\delta$ =0.00). FD-mass spectra were recorded on a Hitachi DF/GC/MS M-80 spectrometer. TLC was performed on precoated glass sheets of silica gel 60 F-254 (E. Merck). Chromatography columns were prepared with silica

gel 60 (70—230 mesh, E. Merck). All reagents were obtained from commercial suppliers and used as received unless otherwise indicated. Dichloromethane was dried over molecular sieves 4A.

Acetolysis of threo-2-(2,4-Difluorophenyl)-2-[1-(methylthio)ethylloxirane (1a): To a solution of threo-2-(2,4-difluorophenyl)-2-[1-(methylthio)ethyl]oxirane (1a) (230.0 mg, 1.00 mmol) in dichloromethane (3.0 ml) was added acetic acid (0.10 ml, 1.7 equiv), and the mixture was stirred at 40  $^{\circ}\mathrm{C}$  for 20 h. The above reaction mixture was diluted with 50 ml of dichloromethane and washed with 20 ml of saturated sodium hydrogencarbonate. The organic layer was dried over anhydrous sodium sulfate, and then filtered and evaporated in vacuo. The residue was purified by column chromatography on 10 g of silica gel and eluted with hexane and ethyl acetate (4:1) to give threo-1acetoxy-2-(2,4-difluorophenyl)-3-methylthio-2-butanol (4a, 230.6 mg, 0.930 mmol, 93.0% yield,  $R_f = 0.5$  developed with a 1:1 mixture of hexane and ether) as a colorless oil, and to give a 1:2 mixture of threo-3-acetoxy-2-(2,4-difluorophenyl)-2-methylthio-1-butanol (5a) and threo-4-acetoxy-3-(2,4difluorophenyl)-3-methylthio-2-butanol (6a) (13.9 mg as a mixture, 0.056 mmol, 5.6% yield,  $R_f = 0.3$  developed with a 1:1 mixture of hexane and ether) as a colorless oil.

4a: IR (neat) 3470, 1740, 1615, 1595, and 1500 cm<sup>-1</sup>; FD-MS m/z (%) 290 (M<sup>+</sup>, 100); <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$ =1.09 (3H, d, J=6.9 Hz, CHCH<sub>3</sub>), 1.88 (3H, s, SCH<sub>3</sub>), 2.21 (3H, s, COCH<sub>3</sub>), 3.22 (1H, q, J=6.9 Hz, CHCH<sub>3</sub>), 3.69 (1H, s, OH), 4.60 (1H, d, J=12.0 Hz, CH<sub>2</sub>), 4.97 (1H, d, J=12.0 Hz, CH<sub>2</sub>), 6.73—6.93 (2H, m), and 7.62 (1H, m).

**5a:** <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  = 1.25 (3H, d, J = 6.9 Hz, CH<u>CH<sub>3</sub></u>), 1.87 (3H, s, SCH<sub>3</sub>), 2.00 (3H, s, COCH<sub>3</sub>), 2.68 (1H, dd, J = 4.0 and 9.2 Hz, OH), 4.06 (1H, dd, J = 4.0 and 12.2 Hz, CH<sub>2</sub>), 4.30 (1H, dd, J = 9.2 and 12.2 Hz, CH<sub>2</sub>), 5.57 (1H, q, J = 6.9 Hz, <u>CH</u>CH<sub>3</sub>), 6.76—6.96 (2H, m), and 7.81 (1H, m).

**6a:** <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  = 1.22 (3H, d, J = 7.0 Hz, CH<u>CH<sub>3</sub></u>), 1.97 (3H, s, SCH<sub>3</sub>), 2.03 (3H, s, COCH<sub>3</sub>), 2.29 (1H, d, J = 7.0 Hz, OH), 4.39 (1H, quint., J = 7.0 Hz, <u>CH</u>CH<sub>3</sub>), 4.74 (1H, d, J = 12.2 Hz, CH<sub>2</sub>), 4.85 (1H, d, J = 12.2 Hz, CH<sub>2</sub>), 6.76—6.96 (2H, m), and 7.71 (1H, m).

Acetolysis of threo- and erythro-2-Aryl-2-[1-(methylthio)ethyl]oxiranes: The following compounds were obtained by the same method as that described for the acetolysis of threo-oxirane 1a. Yields and product ratios are shown in Tables 1 and 2 respectively.

**4b:** <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  = 1.15 (3H, d, J = 7.2 Hz, CH<u>CH<sub>3</sub></u>), 1.96 (3H, s, SCH<sub>3</sub>), 2.08 (3H, s, COCH<sub>3</sub>), 3.06 (1H, q, J = 7.2 Hz, <u>CH</u>CH<sub>3</sub>), 3.16 (1H, s, OH), 4.52 (1H, d, J = 12.6 Hz, CH<sub>2</sub>), 4.58 (1H, d, J = 12.6 Hz, CH<sub>2</sub>), 7.04 (2H, t, J = 8.9 Hz), and 7.41 (2H, dd, J = 5.3 and 8.9 Hz).

**5b:** <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  = 1.16 (3H, d, J = 6.3 Hz, CH<u>CH<sub>3</sub></u>), 1.86 (3H, s, SCH<sub>3</sub>), 2.03 (3H, s, COCH<sub>3</sub>), 2.71 (1H, dd, J = 4.0 and 9.0 Hz, OH), 3.97 (1H, dd, J = 4.0 and 12.0 Hz, CH<sub>2</sub>), 4.10 (1H, dd, J = 9.0 and 12.0 Hz, CH<sub>2</sub>), 5.41 (1H, q, J = 6.3 Hz, <u>CH</u>CH<sub>3</sub>), 6.91—7.11 (2H, m), and 7.30—7.60 (2H, m).

**6b:** <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  = 1.22 (3H, d, J = 6.6 Hz, CH<u>CH</u><sub>3</sub>), 1.82 (3H, s, SCH<sub>3</sub>), 2.08 (3H, s, COCH<sub>3</sub>), 2.24 (1H, d, J = 5.6 Hz, OH), 4.19 (1H, m, <u>CH</u>CH<sub>3</sub>), 4.60 (1H, d, J = 12.0 Hz, CH<sub>2</sub>), 4.83 (1H, d, J = 12.0 Hz, CH<sub>2</sub>), 6.91—7.11 (2H, m), and 7.30—7.60 (2H, m).

**4c:** <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$ =1.16 (3H, d, J=7.0 Hz, CH<u>CH<sub>3</sub></u>), 1.95 (3H, s, SCH<sub>3</sub>), 2.06 (3H, s, COCH<sub>3</sub>), 3.10 (1H, q, J=7.0 Hz, <u>CH</u>CH<sub>3</sub>), 3.15 (1H, s, OH), 4.54 (1H, d, J=11.5 Hz, CH<sub>2</sub>), 4.62 (1H, d, J=11.5 Hz, CH<sub>2</sub>), and 7.24—7.45 (5H, m).

**5c:** <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$ =1.15 (3H, d, J=6.4 Hz, CH<u>CH<sub>3</sub></u>), 1.85 (3H, s, SCH<sub>3</sub>), 2.02 (3H, s, COCH<sub>3</sub>), 2.71 (1H, dd, J=3.6 and 8.9 Hz, OH), 3.99 (1H, dd, J=3.6 and 11.9 Hz, CH<sub>2</sub>), 4.17 (1H, dd, J=8.9 and 11.6 Hz, CH<sub>2</sub>), 5.45 (1H, q, J=6.4 Hz, <u>CH</u>CH<sub>3</sub>), and 7.20—7.60 (5H, m).

**6c:** <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  = 1.24 (3H, d, J = 6.6 Hz, CH<u>CH</u><sub>3</sub>), 1.81 (3H, s, SCH<sub>3</sub>), 2.08 (3H, s, COCH<sub>3</sub>), 2.22 (1H, d, J = 5.6 Hz, OH), 4.23 (1H, m, <u>CH</u>CH<sub>3</sub>), 4.64 (1H, d, J = 11.9 Hz, CH<sub>2</sub>), 4.87 (1H, d, J = 11.9 Hz, CH<sub>2</sub>), and 7.20—7.60 (5H, m).

4d: <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$ =1.17 (3H, d, J=7.3 Hz, CH<u>CH<sub>3</sub></u>), 1.96 (3H, s, SCH<sub>3</sub>), 2.05 (3H, s, COCH<sub>3</sub>), 2.34 (3H, s, PhCH<sub>3</sub>), 3.08 (1H, q, J=7.3 Hz, <u>CH</u>CH<sub>3</sub>), 3.11 (1H, s, OH), 4.52 (1H, d, J=11.5 Hz, CH<sub>2</sub>), 4.59 (1H, d, J=11.5 Hz, CH<sub>2</sub>), 7.15 (1H, d, J=8.0 Hz), and 7.30 (2H, d, J=8.0 Hz).

**5d:** <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  = 1.16 (3H, d, J = 6.3 Hz, CH<u>CH</u><sub>3</sub>), 1.82 (3H, s, SCH<sub>3</sub>), 2.06 (3H, s, COCH<sub>3</sub>), 2.34 (3H, s, PhCH<sub>3</sub>), 2.54 (1H, dd, J=5.3 and 7.6 Hz, OH), 3.88 (1H, dd, J=5.3 and 11.5 Hz, CH<sub>2</sub>), 4.16 (1H, dd, J=7.6 and 11.5 Hz, CH<sub>2</sub>), 5.40 (1H, q, J=6.3 Hz, <u>CH</u>CH<sub>3</sub>), 7.18 (2H, d, J=8.0 Hz), and 7.40 (2H, d, J=8.0 Hz).

**6d:** <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  = 1.13 (3H, d, J = 6.6 Hz, CH<u>CH</u><sub>3</sub>), 1.80 (3H, s, SCH<sub>3</sub>), 2.11 (3H, s, COCH<sub>3</sub>), 2.22 (1H, d, J = 7.0 Hz, OH), 2.34 (3H, s, PhCH<sub>3</sub>), 4.22 (1H, quint., J = 7.0 Hz, <u>CH</u>CH<sub>3</sub>), 4.72 (1H, d, J = 11.9 Hz, CH<sub>2</sub>), 4.82 (1H, d, J = 11.9 Hz, CH<sub>2</sub>), 7.18 (2H, d, J = 8.0 Hz), and 7.40 (2H, d, J = 8.0 Hz).

7a: IR (CHCl<sub>3</sub>) cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$ =1.42 (3H, d, J=7.0 Hz, CHCH<sub>3</sub>), 1.77 (3H, s, SCH<sub>3</sub>), 1.89 (3H, s, COCH<sub>3</sub>), 3.36 (1H, q, J=7.0 Hz, CHCH<sub>3</sub>), 3.60 (1H, s, OH) 4.46 (1H, d, J=11.5 Hz, CH<sub>2</sub>), 4.64 (1H, d, J=11.5 Hz, CH<sub>2</sub>), 6.77 (1H, m), 6.92 (1H, m), and 7.65 (1H, m).

8a: <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  = 1.17 (3H, d, J = 6.6 Hz, CH<u>CH</u><sub>3</sub>), 1.89 (3H, s, SCH<sub>3</sub>), 2.04 (3H, s, COCH<sub>3</sub>), 2.64 (1H, t, J = 7.0 Hz, OH), 4.01 (1H, dd, J = 7.0 and 12.0 Hz, CH<sub>2</sub>), 4.35 (1H, dd, J = 7.0 and 12.0 Hz, CH<sub>2</sub>), 5.52 (1H, q, J = 6.6 Hz, CHCH<sub>3</sub>), 6.78—6.97 (2H, m), and 7.81 (1H, m).

9a:  ${}^{1}$ H NMR (CDCl<sub>3</sub>)  $\delta$ =1.20 (3H, d, J=6.6 Hz, CH<u>CH</u><sub>3</sub>), 1.86 (3H, s, SCH<sub>3</sub>), 2.05 (3H, s, COCH<sub>3</sub>), 2.44 (1H, d, J=5.6 Hz, OH), 4.39 (1H, m, <u>CH</u>CH<sub>3</sub>), 4.77 (1H, d, J=12.0 Hz, CH<sub>2</sub>), 4.86 (1H, d, J=12.0 Hz, CH<sub>2</sub>), 6.78—6.97 (2H, m), and 7.56 (1H, m).

7b: <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta = 1.27$  (3H, d, J = 7.2 Hz, CH<u>CH<sub>3</sub></u>), 1.92 (3H, s, SCH<sub>3</sub>), 2.00 (3H, s, COCH<sub>3</sub>), 3.07 (1H, q, J = 7.2 Hz, <u>CH</u>CH<sub>3</sub>), 3.21 (1H, s, OH), 4.47 (1H, d, J = 11.6 Hz, CH<sub>2</sub>), 4.62 (1H, d, J = 11.6 Hz, CH<sub>2</sub>), 7.04 (2H, t, J = 8.6 Hz), and 7.48 (2H, dd, J = 5.3 and 8.6 Hz).

8b: <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$ =1.16 (3H, d, J=6.6 Hz, CH<u>CH</u><sub>3</sub>), 1.82 (3H, s, SCH<sub>3</sub>), 2.06 (3H, s, COCH<sub>3</sub>), 2.52 (1H, dd, J=5.6 and 7.9 Hz, OH), 3.87 (1H, dd, J=5.6 and 11.9 Hz, CH<sub>2</sub>), 4.14 (1H, dd, J=7.9 and 11.9 Hz, CH<sub>2</sub>), 5.38 (1H, q, J=6.6 Hz, <u>CH</u>CH<sub>3</sub>), 7.08 (2H, t, J=8.9 Hz), and 7.53 (2H, dd, J=5.3 and 8.9 Hz).

9b: <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$ =1.12 (3H, d, J=6.6 Hz, CH<u>CH</u><sub>3</sub>), 1.83 (3H, s, SCH<sub>3</sub>), 2.11 (3H, s, COCH<sub>3</sub>), 2.28 (1H, d, J=6.6 Hz, OH), 4.20 (1H, quint., J=6.6 Hz,

 $\underline{\text{CH}}\text{CH}_3$ ), 4.67 (1H, d, J=11.9 Hz,  $\text{CH}_2$ ), 4.83 (1H, d, J=11.9 Hz,  $\text{CH}_2$ ), 7.07 (2H, t, J=9.2 Hz), and 7.53 (2H, dd, J=5.3 and 8.9 Hz).

7c: <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$ =1.28 (3H, d, J=7.2 Hz, CH<u>CH</u><sub>3</sub>), 1.89 (3H, s, SCH<sub>3</sub>), 1.99 (3H, s, COCH<sub>3</sub>), 3.11 (1H, q, J=7.2 Hz, <u>CH</u>CH<sub>3</sub>), 3.19 (1H, s, OH), 4.49 (1H, d, J=11.5 Hz, CH<sub>2</sub>), 4.65 (1H, d, J=11.5 Hz, CH<sub>2</sub>), and 7.28—7.52 (5H, m).

8c: <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta = 1.17$  (3H, d, J = 6.3 Hz, CH<u>CH<sub>3</sub></u>), 1.82 (3H, s, SCH<sub>3</sub>), 2.06 (3H, s, COCH<sub>3</sub>), 2.55 (1H, dd, J = 5.6 and 7.6 Hz, OH), 3.91 (1H, dd, J = 5.6 and 11.5 Hz, CH<sub>2</sub>), 4.19 (1H, dd, J = 7.6 and 11.5 Hz, CH<sub>2</sub>), 5.43 (1H, q, J = 6.3 Hz, CHCH<sub>3</sub>), and 7.26—7.56 (5H, m).

**9c:** <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  = 1.13 (3H, d, J = 6.6 Hz, CH<u>CH</u><sub>3</sub>), 1.81 (3H, s, SCH<sub>3</sub>), 2.11 (3H, s, COCH<sub>3</sub>), 2.24 (1H, d, J = 6.6 Hz, OH), 4.24 (1H, quint., J = 6.6 Hz, C<u>H</u>CH<sub>3</sub>), 4.74 (1H, d, J = 11.9 Hz, CH<sub>2</sub>), 4.85 (1H, d, J = 11.9 Hz, CH<sub>2</sub>), and 7.28—7.55 (5H, m).

7d: <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  = 1.27 (3H, d, J = 7.0 Hz, CH<u>CH<sub>3</sub></u>), 1.92 (3H, s, SCH<sub>3</sub>), 2.00 (3H, s, COCH<sub>3</sub>), 2.34 (3H, s, PhCH<sub>3</sub>), 3.09 (1H, q, J = 7.0 Hz, <u>CH</u>CH<sub>3</sub>), 3.14 (1H, s, OH), 4.48 (1H, d, J = 11.5 Hz, CH<sub>2</sub>), 4.62 (1H, d, J = 11.5 Hz, CH<sub>2</sub>), 7.16 (2H, d, J = 8.0 Hz), and 7.37 (2H, d, J = 8.0 Hz).

8d:  $^{1}$ H NMR (CDCl<sub>3</sub>)  $\delta$ =1.16 (3H, d, J=6.6 Hz, CH<u>CH<sub>3</sub></u>), 1.82 (3H, s, SCH<sub>3</sub>), 2.06 (3H, s, COCH<sub>3</sub>), 2.35 (3H, s, PhCH<sub>3</sub>), 2.53 (1H, dd, J=5.6 and 7.6 Hz, OH), 3.88 (1H, dd, J=5.6 and 11.5 Hz, CH<sub>2</sub>), 4.16 (1H, dd, J=7.6 and 11.5 Hz, CH<sub>2</sub>), 5.40 (1H, q, J=6.6 Hz, CHCH<sub>3</sub>), 7.18 (2H, d, J=8.0 Hz), and 7.40 (2H, d, J=8.0 Hz).

**9d:** <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  = 1.13 (3H, d, J = 6.6 Hz, CH<u>CH</u><sub>3</sub>), 1.80 (3H, s, SCH<sub>3</sub>), 2.11 (3H, s, COCH<sub>3</sub>), 2.21 (1H, d, J = 7.0 Hz, OH), 2.35 (3H, s, PhCH<sub>3</sub>), 4.22 (1H, quint., J = 7.0 Hz, <u>CH</u>CH<sub>3</sub>), 4.72 (1H, d, J = 11.9 Hz, CH<sub>2</sub>), 4.82 (1H, d, J = 11.9 Hz, CH<sub>2</sub>), 7.18 (2H, d, J = 8.0 Hz), and 7.40 (2H, d, J = 8.0 Hz).

erythro-2-(2,4-Difluorophenyl)-2-methylthio-1,3butanediol (12a): To a solution of erythro-3-acetoxy-2-(2,4-difluorophenyl)-2-methylthio-1-butanol (8a) and erythro-4-acetoxy-3-(2,4-difluorophenyl)-3-methylthio-2-butanol (9a) (152.0 mg, 0.524 mmol, 8a/9a=3:1) in methanol (1.5 ml) was added anhydrous potassium carbonate (150 mg, 2.0 equiv), and the mixture was stirred at room temperature for 3 h. Water (20 ml) was added to the reaction mixture, which was then extracted with chloroform (30 ml×3). The combined organic layers were washed with 30 ml of saturated ammonium chloride, dried over anhydrous sodium sulfate, filtered, and evaporated in vacuo to give diol 12a (120.7 mg, 0.487 mmol, 92.9% yield: Colorless crystals; mp 62.5—64.5 °C; IR (KBr) 3400, 1610, 1590, and 1500 cm<sup>-1</sup>; FD-MS m/z (%) 248 (M<sup>+</sup>, 100); <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$ =1.18 (3H, d, J=6.9 Hz, CH<u>CH</u><sub>3</sub>), 1.77 (3H, s, SCH<sub>3</sub>), 2.77 (1H, d, J=8.0 Hz, OH), 2.94 (1H, dd, J=4.3 and 8.9 Hz, OH), 4.05 $(1H, dd, J=4.3 \text{ and } 12.0 \text{ Hz}, CH_2), 4.37 (1H, dq, J=8.0 \text{ and})$ 6.9 Hz, CHCH<sub>3</sub>), 4.56 (1H, dd, J=8.9 and 12.0 Hz, CH<sub>2</sub>), 6.77—6.95 (2H, m), and 7.56 (1H, m). Found: C, 53.20; H, 5.71; S, 13.24%. Calcd. for  $C_{11}H_{14}O_2F_2S$ : C, 53.21; H, 5.68; S, 12.91%.

erythro-2-(2,4-Difluorophenyl)-2-methylsulfonyl-1,2-butanediol (13): To a solution of diol 12a (189.6 mg, 0.764 mmol) in dichloromethane (3.0 ml) was added 4-chloroperbenzoic acid (494 mg, 80% assay, 3.0 equiv) at 0

°C. After being stirred at 0 °C for 30 min, the mixture was stirred at room temperature overnight. The reaction mixture was quenched with 20 ml of 10% aq Na<sub>2</sub>SO<sub>3</sub> and diluted with 100 ml of ethyl acetate. The mixture was stirred for 5 min, and the aqueous layer was then separated and extracted with 50 ml of ethyl acetate. The combined organic layers were washed with 30 ml of saturated sodium hydrogencarbonate, dried over anhydrous sodium sulfate, filtered, and evaporated in vacuo to give 223 mg of sulfone 13 as a crude crystalline powder. This was recrystallized from dichloromethane to give pure 13 as colorless crystals (174.7 mg, 0.623 mmol, 81.6% yield: mp 128.5—129.0 °C; IR (KBr) 3400, 1615, 1600, and 1510 cm<sup>-1</sup>; <sup>1</sup>H NMR (CD<sub>3</sub>OD)  $\delta = 1.14$  (3H, d, J = 6.9 Hz, CHCH<sub>3</sub>), 3.03 (3H, s, SO<sub>2</sub>CH<sub>3</sub>), 4.67 (2H, s, CH<sub>2</sub>), 5.16 (1H, q, J=6.9 Hz, <u>CH</u>CH<sub>3</sub>), 6.92-7.06 (2H, m), and 7.85 (1H, m). Found: C, 46.97; H, 5.07; S, 11.80%. Calcd for C<sub>11</sub>H<sub>14</sub>O<sub>4</sub>F<sub>2</sub>S: C, 47.14; H, 5.03; S, 11.44%.

Preparation of threo-Epoxide 1a Free from Its erythro-Isomer 2a: To a solution of a threo-rich mixture of oxiranes 1a and 2a (300.0 mg, 1.303 mmol) in dichloromethane (3.0 ml) was added acetic acid (0.10 ml, 1.35 equiv), and the mixture was stirred at 40 °C for 16 h. This reaction mixture was diluted with 50 ml of dichloromethane and washed with 20 ml of saturated sodium hydrogencarbonate. The organic layer was dried over anhydrous sodium sulfate, and then filtered and evaporated in vacuo. The residue was purified by column chromatography on 15 g of silica gel being eluted with hexane and ethyl acetate (4:1) to give threo-1-acetoxy-2-(2,4-difluorophenyl)-3-methylthio-2-butanol (4a, 265.3 mg, 0.915 mmol, 70.2% yield). The above obtained 4a (97.6 mg, 0.336 mmol) was dissolved in 1.0 ml of methanol. To the solution was added anhydrous potassium carbonate (100 mg, 2.0 equiv), and the mixture was stirred at room temperature for 3 h. Water (20 ml) was added to the reaction mixture, which was then extracted with chloroform (30 ml×3). The combined organic layers were washed with 30 ml of saturated ammonium chloride, dried over anhydrous sodium sulfate, filtered, and evaporated in vacuo to give threo-diol 14a (74.0 mg, 0.298 mmol, 88.8% yield): A colorless oil; IR (neat) 3350, 1610, 1595, and 1500 cm<sup>-1</sup>; FD-MS m/z (%) 290 (M<sup>+</sup>, 100); <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$ =1.12 (3H, d, J=6.9 Hz, CHCH<sub>3</sub>), 2.00 (1H, br, OH), 2.20 (3H, s, SCH<sub>3</sub>), 3.26 (1H, q, J=6.9 Hz,  $\underline{\text{CH}}$ CH<sub>3</sub>), 3.55 (1H, br, OH), 3.83 (1H, d, J=11.5 Hz, CH<sub>2</sub>), 4.32 (1H, d, J=11.5 Hz, CH<sub>2</sub>), 6.72-7.02 (2H, m), and 7.70(1H, m). Found: C, 53.15; H, 5.56; S, 12.90%. Calcd for  $C_{11}H_{14}O_4F_2S$ : C, 53.21; H, 5.68; S, 12.91%.

Diol 14a (50.6 mg, 0.204 mmol) was then dissolved in 2.0 ml of dry dichloromethane under argon atmosphere. To the solution was added triethylamine (0.1 ml, 3.6 equiv), and the mixture was cooled to 0 °C. Methanesulfonyl chloride was added dropwise to the mixture over a 5 min period, and the mixture was stirred at 0 °C. After 30 min, 15% potassium hydroxide solution in water (0.30 g, 4.0 equiv) was added to the reaction mixture, and the resultant two-phase solution was stirred vigorously at 0 °C for 3 h. Water (10 ml) was added to the mixture, which was then extracted with dichloromethane (20 ml×2). The combined organic layers were washed with water (30 ml×3), dried over anhydrous sodium sulfate, filtered, and evaporated in vacuo to give threo-epoxide 1a in a quantitative yield (47.1 mg, 0.204 mmol). The

ratio of 1a vs. 2a was determined to be 360:1 by HPLC.<sup>5)</sup>

Preparation of threo-Epoxides 1b—d Free from Their erythro-Isomers 2b—d: threo-Epoxides 1b—d free from their erythro-isomers 2b—d were obtained by the same method as that described in the preparation of threo-oxirane 1a. The <sup>1</sup>H NMR spectra of intermediate diols 14b—d are shown below.

14b: <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$ =1.18 (3H, d, J=6.9 Hz, CH<u>CH<sub>3</sub></u>), 2.09 (3H, s, SCH<sub>3</sub>), 2.50 (1H, t, J=6.8 Hz, OH), 3.11 (1H, q, J=6.9 Hz, <u>CH</u>CH<sub>3</sub>), 3.41 (1H, s, OH), 3.78 (1H, dd, J=6.8 and 11.5 Hz, CH<sub>2</sub>), 4.06 (1H, dd, J=6.8 and 11.5 Hz, CH<sub>2</sub>), 7.04 (2H, t, J=8.9 Hz), and 7.70 (2H, dd, J=5.3 and 8.9 Hz).

14c: <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$ =1.19 (3H, d, J=7.2 Hz, CH<u>CH</u><sub>3</sub>), 2.09 (3H, s, SCH<sub>3</sub>), 2.43 (1H, t, J=6.9 Hz, OH), 3.14 (1H, q, J=7.2 Hz, <u>CH</u>CH<sub>3</sub>), 3.38 (1H, s, OH), 3.81 (1H, dd, J=6.9, 11.5 Hz, CH<sub>2</sub>), 4.11 (1H, dd, J=6.9, 11.5 Hz, CH<sub>2</sub>), and 7.25—7.45 (5H, m).

**14d:** <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$ =1.20 (3H, d, J=7.0 Hz, CH<u>CH</u><sub>3</sub>), 2.08 (3H, s, SCH<sub>3</sub>), 2.34 (3H, s, PhCH<sub>3</sub>), 2.36 (1H, dd, J=6.3 and 7.6 Hz, OH), 3.12 (1H, q, J=7.0 Hz, <u>CH</u>CH<sub>3</sub>), 3.35 (1H, s, OH), 3.80 (1H, dd, J=6.3 and 11.6 Hz, CH<sub>2</sub>), 4.08 (1H, dd, J=7.6 and 11.6 Hz, CH<sub>2</sub>), 7.17 (2H, d, J=8.0 Hz), and 7.70 (2H, d, J=8.0 Hz).

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### References

- 1) a) I. Saji, K. Tamoto, T. Tanio, T. Okuda, and T. Atsumi, "8th Medicinal Chemistry Symposium," Osaka, Nov. 1986, Abstr., pp. 9—12; b) I. Saji, Y. Tanaka, K. Ichise, T. Tanio, T. Okuda, and T. Atsumi, European Patent 178533 (1986); Chem. Abstr., 105, 35613r (1986).
- 2) I. Saji, K. Tamoto, Y. Tanaka, H. Miyauchi, K. Fujimoto, and N. Ohashi, *Bull. Chem. Soc. Jpn.*, **67**, 1427 (1994).
- 3) a) A. Tasaka, N. Tamura, Y. Matusita, K. Teranishi, R. Hayashi, K. Okonogi, and K. Itoh, *Chem. Pharm. Bull.*, 41, 1035 (1993); b) T. Konosu, T. Miyaoka, Y. Tajima, and S. Oida, *Chem. Pharm. Bull.*, 39, 2241 (1991); c) M. Ogata, H. Matsumoto, S. Shimizu, S. Kida, M. Shiro, and K. Tawara, *Eur. J. Med. Chem.*, 24, 137 (1989).
- 4) K. Yamakawa, K. Nishitani, and K. Kasahara, *Chem. Pharm. Bull.*, **27**, 953 (1979), and references cited therein.
- 5) The ratio of threo-/erythro-epoxides was determined by HPLC. HPLC conditions are as follows: Column; Sumipax ODS A-212 (5  $\mu$ m, 6 mm $\phi \times 25$  cm), mobile phase; acetonitrile/water=55/45, flow rate; 1.0 ml min<sup>-1</sup>, detection; UV 210 nm: In the case of 1a/2a, the peak at 17.4 min. corresponded to threo-epoxide 1a, and the peak at 16.3 min. corresponded to erythro-one 2a.
- 6) M. Ladika, B. Jursic, Z. Mihalic, and D. E. Sanko, *Tetrahedron Lett.*, **27**, 1703 (1986), and references cited therein
- 7) a) A. R. Derzhinskii, L. D. Konyushkin, and A. I. Lutsenko, *Izv. Akad. Nauk SSSR*, 11, 2652 (1984); b) D. M. Gill, N. A. Pegg, and C. M. Rayner, *J. Chem. Soc.*, *Perkin Trans. 1*, 1993, 1371.

8) Y. Tanaka, T. Yuasa, Y. Kawakami, K. Terashima, T. Morita, A. Nishikawa, K. Bando, and M. Kawashima,

European Patent 552974 (1993); Chem. Abstr., 120, 134499 (1993).